



Cytochrome 2C19 Enzyme Polymorphism Frequency in Different Indigenous Ethnic Groups in Russian Federation: A Systematic Review

Karin Mirzaev^{1*}, Dmitriy Sychev², Goar Arutyunyan¹, Alla Yugay¹ and Denis Andreev³

¹I.M. Sechenov First Moscow State Medical University, Moscow, Russian Federation

²Department of Clinical Pharmacology, Russian Medical Academy of Postgraduate Education, Moscow, Russian Federation

³Department of Preventative Cardiology and Acute Cardiac Care, I.M. Sechenov First Moscow State Medical University, Moscow, Russian Federation

Abstract

Background and objective: Genetically determined diversity in the activity of cytochrome P450 (CYP) – enzyme regulating the biotransformation of drugs and xenobiotics – is one of the main causes of interindividual differences in response to pharmacotherapy. The objective of this review is to analyze the prevalence of polymorphic markers of gene *CYP2C19*, associated with the violation of the pharmacological response to clopidogrel among the various ethnic groups living in the Russian Federation.

Methods: A literature review was conducted using the following databases: MEDLINE and eLIBRARY.RU. Russian language articles published between 2003-2003 (the first publication in Russians) and 2014 were reviewed.

Results: The authors detected 11 original research studies on *CYP2C19* gene in 11 indigenous ethnic groups of Russian Federation. According to the research data, the frequency of *CYP2C19*2* and *CYP2C19*3* markers prevalence is higher in the Mongolian race (with maximum *CYP2C19*2* frequency in the Kalmyks – 25, 0 % and *CYP2C19*3* in Tatars – 21,0 %). *CYP2C19*17* allele has been studied only in Russians, and was about the same as in the Caucasian race (14,0 %).

Conclusion: The results of the investigation will be beneficial for developing guidelines for *CYP2C19* genotype-directed antiplatelet therapy for each region of Russia.

Keywords: *CYP2C19* Polymorphism; Pharmacogenetics; Ethnic differences of *CYP2C19*2*; *CYP2C19*3*; *CYP2C19*17*; Clopidogrel resistance

Introduction

Individual's response to specific drugs is a great issue for medicine in the twenty-first century. Genetically determined diversity in the activity of cytochrome P450 (CYP) – enzyme regulating the biotransformation of drugs and xenobiotics – is one of the main causes of the interindividual differences in response to pharmacotherapy.

Cytochrome P450 was first described in 1958 by Klingenberg [1] and Garfinkel [2]. The known clinically relevant cytochromes include CYP1A2, CYP2C9, CYP2C19, CYP2D6, CYP2E1 и CYP3A4. Some of these isoforms exhibit genetic polymorphisms. The frequency of these polymorphisms differs markedly between ethnic groups. These genetic differences mean some people have an enzyme with reduced or no activity. Patients who are 'slow metabolisers' may have an increased risk of adverse reactions to a drug metabolised by the affected enzyme. It is estimated that genetics can account for 20 to 50 percent of variability in drug disposition and effects [3].

CYP2C19 appears to be one of the main CYP2C isoform found in the human. CYP2C19 hydroxylates a wide a wide variety of drugs (clopidogrel, barbiturates, diazepam, lansoprazole, nelfinavir, clonazepam, cyclophosphamide, omeprazole, etc.) [4].

Genetic polymorphism was discovered at Vanderbilt University by Kupfer et al. [5] in 1979 when conducting the research 4'-hydroxylation of the anticonvulsant S-mephenytoin. Later in 1993 Wrington et al. [6] found out that S-mefenitoin was the substrate of CYP2C19 enzyme. In 1994, Goldstein and de Morais [7] found that *CYP2C19* gene polymorphisms are associated with the loss of heterozygosity on chromosomes 10q (10q.1-24.3).

There is about 34 *CYP2C19* alleles including *CYP2C19*1*, *CYP2C19*2* and *CYP2C19*3*, *CYP2C19*17* (<http://www.cypalleles.ki.se/cyp2c19.htm>).

Among functional defective alleles *CYP2C19*2* contributes 75% [8] in Asians and 93% [9] in Caucasians. 25% of defective alleles in Asians [10] is the *CYP2C19*3*, which is very rare in Caucasians (less than 1%) [11]. These pharmacogenetic variations lead to inappropriate concentrations of drugs and drug metabolites, which may contribute towards the toxicity and risk of adverse drug reactions or lack of therapeutic benefit. In contrast, the pro-drugs such as clopidogrel may be less effective in reducing the rate of cardiovascular events among persons who are carriers of loss-of-function CYP2C19 alleles that are associated with reduced conversion of clopidogrel to its active metabolite.

Several studies have examined the frequency of various *CYP2C19* alleles worldwide. The reported allele frequency of *CYP2C9*2* was about 50% in Asians, 18% in Caucasians, 34% in Africans and 19% in American populations [12-15]. The allele frequency of *CYP2C9*3* among the Caucasian, African and Asian populations was <1 %, <1 % and 7 %, respectively [16]. The *CYP2C19*17* genotype was found in 25,7 % of the Germans [17], 22,0 % of the Norse [18], 20,0 % of the Swedes [19], 0,3 % of the Koreans [19], 4,0 % of the Chinese [20], 1,3 % of the Japanese [21].

***Corresponding author:** Karin Mirzaev, I.M. Sechenov First Moscow State Medical University, Moscow, Russian Federation, Trubetskaya 8 street, 119991 Moscow, Russian Federation, Tel: +79852720253; Fax: +7 495 9155801; E-mail: karin05doc@yandex.ru

Received April 24, 2014; Accepted June 27, 2014; Published July 04, 2014

Citation: Mirzaev K, Sychev D, Arutyunyan G, Yugay A, Andreev D (2014) Cytochrome 2C19 Enzyme Polymorphism Frequency in Different Indigenous Ethnic Groups in Russian Federation: A Systematic Review J Pharmacogenomics Pharmacoproteomics 5: 136. doi:10.4172/2153-0645.1000136

Copyright: © 2014 Mirzaev K, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Ethnic groups	Region Of Russian Federation	n	Polymorphic marker	Population characteristics	Genotypes n (%)				Allelic variants				References
					EM	IM	PM	UP	*1	*2	*3	*17	
Russians	Tomsk	130	*2	Allergic diseases	81 (66,9)	38(31,4)	2 (1,7)	n.d.	0,83	0,17	n.d.	n.d.	[28]
	Tomsk	62	*2,*3	lymphoproliferative disorders	n.d. ¹	n.d.	n.d.	n.d.	0,86	0,14	0,0	n.d.	[37]
	Moscow and the Moscow Region	395	*2	Ischemic heart disease	288 (72,7)	101(25,5)	7 (1,8)	n.d.	0,85	0,15	n.d.	n.d.	[36]
	Moscow and the Moscow Region	40	*2,*3,*17	Ischemic heart disease	20 (50,0)	12 (30,0)	0 (0,0)	8 (20,0)	0,71	0,15	0,0	0,14	[35]
		146	*2	Healthy	111 (76,1)	31 (21,2)	4 (2,7)	n.d.	0,83	0,13	n.d.	n.d.	
	Tomsk	82	*2	Healthy	64 (78,0)	16 (19,5)	0 (0,0)	n.d.	0,88	0,12	n.d.	n.d.	[34] ²
		87	*3	Healthy	87 (100,0)	0 (0,0)	0 (0,0)	n.d.	1	n.d.	0,0	n.d.	
	Astrakhan	52	*2	Healthy	40(77,0)	8 (16,0)	4(7,0)	n.d.	0,85	0,15	n.d.	n.d.	[30]
Voronezh	290	*2,*3	Healthy	228 (78,7)	56 (19,3)	6 (2,0)	n.d.	0,86	0,11	0,03	n.d.	[38]	
Tatars	Kazan	97	*2	Ischemic heart disease	76 (78,4)	20 (20,6)	1 (1,0)	n.d.	0,86	0,14	n.d.	n.d.	[39]
	The Republic of Tatarstan	130	*2,*3	Acid-related disorders	56 (43,1)	62 (47,7)	12 (9,2)	n.d.	0,67	0,12	0,21	n.d.	[40]
	Astrakhan	50	*2	Healthy	40 (80,0)	7 (14,0)	3 (6,0)	n.d.	0,87	0,13	n.d.	n.d.	[30]
Kalmyks	Astrakhan	50	*2	Healthy	31 (62,0)	13 (26,0)	6 (12,0)	n.d.	0,85	0,15	n.d.	n.d.	[30]
Tuvinians	The Republic of Tuva	88	*2	Healthy	63 (71,7)	24 (27,2)	1 (1,1)	n.d.	0,85	0,15	n.d.	n.d.	[34]
		88	*3	Healthy	84 (95,4)	4 (4,6)	0 (0,0)	n.d.	0,98	n.d.	0,02	n.d.	
Buryats	The Republic of Buryatia	88	*2	Healthy	54 (61,3)	31 (35,2)	3 (3,5)	n.d.	0,79	0,21	n.d.	n.d.	[34]
		88	*3	Healthy	77 (87,5)	10 (11,4)	1 (1,1)	n.d.	0,93	n.d.	0,07	n.d.	
Yakuts	The Sakha Republic (Yakutia)	88	*2	Healthy	54 (61,3)	27 (30,6)	7 (7,9)	n.d.	0,77	0,23	n.d.	n.d.	[34]
		87	*3	Healthy	79 (90,8)	8 (9,2)	0 (0,0)	n.d.	0,95	n.d.	0,05	n.d.	
Altayans	Altai Republic	87	*2	Healthy	64 (73,5)	20 (23,0)	3 (3,5)	n.d.	0,85	0,15	n.d.	n.d.	[34]
		87	*3	Healthy	80 (92,0)	7 (8,0)	0 (0,0)	n.d.	0,96	n.d.	0,04	n.d.	
Chechens	Astrakhan	50	*2	Healthy	41(82,0)	7 (14,0)	2 (6,0)	n.d.	0,87	0,13	n.d.	n.d.	[30]
Carachays	The Republic of Karachay-Cherkessia	125	*2	Healthy	92 (73,6)	31 (24,8)	2 (1,6)	n.d.	0,86	0,14	n.d.	n.d.	[31]
Circassians		77	*2	Healthy	53 (68,8)	19 (24,7)	5 (6,5)	n.d.	0,81	0,19	n.d.	n.d.	[31]
Ingushes	Astrakhan	50	*2	Healthy	44 (88,0)	4 (8,0)	2 (4,0)	n.d.	0,92	0,08	n.d.	n.d.	[30]
Dagestans (Laks, Dargins, Avars)	Makhachkala	30	*2	Healthy	26 (86,7)	4 (13,3)	n.d.	n.d.	0,93	0,07	n.d.	n.d.	[32]

Table 1: Allele and genotype frequencies of CYP2C19 gene in different ethnic groups in Russia.

¹n.d. = Not determined for the study population by authors.

²In the study by Makeeva et al. [34] prevalence of CYP2C19*2 and CYP2C19*3 was studied in separate groups

Determining the CYP2C19 genotype can help by determining the metabolizer phenotype. Normal CYP2C19 enzyme activity is expected when two CYP2C19*1 alleles are considered to be present (CYP2C19*1/*2); CYP2C19 Intermediate Metabolizer (IM) phenotype is suggested by the presence of one CYP2C19 allele with decreased function and one CYP2C19 allele with normal function or one CYP2C19 allele with decreased function and one CYP2C19 allele with increased function (CYP2C19*1/*2, *1/*3, *2/*17, *3/*17); CYP2C19 Poor Metabolizer (PM) phenotype is suggested by the presence of two CYP2C19 non-functional alleles CYP2C19*2 or CYP2C19*3 (CYP2C19*2/*2, *2/*3, *3/*3). Heterozygosity or homozygosity for the increased function CYP2C19*17 allele is associated with increased CYP2C19 activity and an ultra-rapid metabolizer phenotype (UM) (<http://www.pharmgkb.org/>).

Approximately 3% of the European population, 4-7% of the African population [4], 12-16 % of the Koreans [22,23], 18-23 % of the Japanese [24,25], 15-17 % of the Chinese [26] are CYP2C19-poor metabolizers.

Extensive and intermediate metabolizers phenotypes are the most common in humans, because CYP2C19 poor-metabolizer phenotypes behave as autosomal recessive traits [8].

Since cytochrome enzymes are responsible for metabolizing over

half of all drugs on the market today, it is important for a physician to have valuable information to determine whether a patient's specific genotype may impact their drug response. Moreover, knowing the CYP2C19 phenotype of a patient may help in prescribing optimum dose of drug to achieve better therapeutic outcome.

The Russian Federation is a geographically huge country with a vast variety of ethnic groups. Nowadays, there is a lack of publications referred to CYP2C19 gene polymorphisms prevalence among the different ethnicities in the Russian Federation (except Russians). Therefore, the aim of this study was: (1) to analyze the prevalence of polymorphic markers of gene CYP2C19 in various ethnic groups living in the Russian Federation and (2) acquaint foreign researchers with the this data.

Materials and Methods

We conducted a systematic literature review to identify published studies of CYP2C19 allelic variations and frequencies for different indigenous ethnic groups in Russian Federation. A literature review was conducted using the following databases: MEDLINE and eLIBRARY. RU. Russian language articles published between 2003 and 2014 were reviewed.

Search terms “Cytochrome P450”, “*CYP2C19*”, “*CYP2C19*2*”, “*CYP2C19*3*”, “*CYP2C19*17*”, “Genetic polymorphism of *CYP2C19*”, “Pharmacogenetics” were used.

Included studies had to meet the following inclusion criteria: (1) *CYP2C19* genotyping performed in all patients, (2) there is an indication of the ethnicity of participants in all studies, (3) original studies published between 2003 (the first publication in Russians) and 2014. Exclusion criteria: review articles.

The following data were abstracted: population characteristics (healthy or patients), number of subjects, ethnicity, frequency of alleles and genotypes, region of population residence.

There were no restrictions of inclusion on the basis of patient characteristics, publication type (journal article, abstract or conference proceedings), or publication language.

Results and Discussion

We detected 11 original research studies on *CYP2C19* gene in 11 indigenous ethnic groups in Russian Federation (Table 1). These data may confer important benefits in terms of determination of appropriate strategies of drug therapy, clinical safety and for best decision-making in public health about the rational use of *CYP2C19* substrates in different indigenous ethnic groups in Russian Federation. However, lack of information about frequency of *CYP2C19* alleles could create a barrier to the use of pharmacogenetic testing in these populations [27].

Ethnic distribution of *CYP2C19* alleles and genotypes was studied among Russians, Tatars, Karachays, Circassians, Ingushes, Chechens, Kalmuks and Dagestan's people (Laks, Dargins, Avars). The ethnicity was identified on the basis of patient's ethnic self-identification. In some cases the researchers surveyed the parents of the trial subjects in order to identify ethnicity.

Freidin's et al. was investigated 130 Russians living in Russian city of Tomsk were enrolled (median age 39 ± 13 , 5 years, 67 women and 63 men) [28]. *CYP2C19*2* allele frequency was 14,7%, *CYP2C19*1*1*, *CYP2C19*1*2* and *CYP2C19*2*2* genotype frequencies – 66,9 % (81 participants), 31,4 % (38 participants) и 1,7 % (2 participants) respectively. The results shown in this work confirmed the data of studies conducted among Caucasians [29], at the same time *CYP2C19*2* frequency is remarkably lower than that in the Mongolian race [8].

Kantemirova B.I. et al. identified *CYP2C19*2* among Russians, Chechens, Tatars, Kalmyks and Ingushes. The study included 208 healthy children aged from 1 to 18 years [30]. According to the study, for functional deficient *CYP2C19*2*, allele frequency is highest among Kalmyks – 25%; 11,0% in Chechens; 14% in Tatars; 8,0% in Ingushes.

The difference between *CYP2C19* genotype (Table 1) frequencies among Kalmyks and Ingushes ($\chi^2=5,765$, $p=0,0163$) as well as between the Kalmyks and the Chechens ($\chi^2=3,6$, $p=0,0289$) were statistically significant. Relatively high allele and genotype *CYP2C19*2* frequencies in Kalmyks are natural, as Kalmyks belong to Mongolian race. The cause of low incidence of the *CYP2C19*2* allelic variant in the research among Tatars is probably mixing with other ethnic groups and incorrect selection of patients.

The polymorphic marker *CYP2C19*2* has also been identified among other indigenous ethnic groups of the North Caucasus: Karachays and Circassians [31], Laks, Dargins, Avars [32].

Romodanovsky et al. [31] investigated 202 participants: 77 Circassians and 125 Karachays (median age 56 ± 11 , 31 men and 46

women). The *CYP2C19*2* allele frequencies among Karachays and Circassians were 18,8 % and 14,0 % respectively.

In Dagestan's peoples (Laks, Dargins, Avars) was observed the lowest rate of *CYP2C19*2* polymorphism in Russian Federation – 6,5 %.

On the whole the *CYP2C19*2* allele frequencies in ethnic groups of the North Caucasus are close to those received earlier among most of the nations of Caucasian (White) race [29], which is natural, as Karachayevs, Cherkesses, Ingushes. Laks, Dargins, Avars belong to Caucasian race (not to be confused “Caucasian” and “Caucasus”!).

The frequency of *CYP2C19*2* polymorphic marker was also studied among Bashkirs [33], Yakuts, Buryats, Altayans and Tuvinians [34] (Table 1). The results are close to those received among the Mongolian race.

In our literature review we have evaluated the prevalence of the *CYP2C19* gene polymorphisms among the 11 ethnicities in the Russian Federation. As it was expected, *CYP2C19*2* allele prevalence was higher among the Asian population, with the highest rate in Kalmyks - 25,0 %, The highest rate of *CYP2C19*3* polymorphism was observed in Tatars - 21,2 %.

However, the high rate of *CYP2C19*3* polymorphism is very uncommon in humans (up to 5-7% in the Asian population and about 1% in the European population [16]) and this phenomenon calls for additional studies. *CYP2C19*17* allele prevalence in the Russian population was observed in one study [35] and it was similar to those in the European population (14,0%).

In general, the *CYP2C19* gene polymorphisms prevalence among the Tatars, Kalmyks, Yakuts, Tuvins, Buryats and Altays was similar to those in mongoloids. The *CYP2C19* gene polymorphisms prevalence among the Russians, Karachayevs, Cherkesses, Ingushes, Laks, Dargins, Avars was similar to those in the European population [36-40].

Conclusion

- The evaluation of the interindividual differences in the prevalence of *CYP2C19* gene polymorphisms is very important in the Russian Federation because of the high multinationality. The results of the pharmacogenetic investigation may be beneficial for developing guidelines for *CYP2C19* genotype-directed antiplatelet therapy for each region of the Russian Federation.
- Since cytochrome enzymes are responsible for metabolizing over half of all drugs on the market today, it is important for a physician to have valuable information to determine whether a patient's specific genotype may impact their drug response. Moreover, knowing the *CYP2C19* phenotype of a patient may help in prescribing optimum dose of drug and in predicting the increased risk of adverse reactions to achieve better therapeutic outcome.

Competing Interests

The authors declare that they have no competing interests.

References

1. Klingenberg M (1958) Pigments of rat liver microsomes. Arch Biochem Biophys 75: 376-386.
2. Garfinkel D (1958) Studies on pig liver microsomes. I. Enzymic and pigment composition of different microsomal fractions. Arch Biochem Biophys 77: 493-509.

3. Evans WE, McLeod HL (2003) Pharmacogenomics--drug disposition, drug targets, and side effects. *N Engl J Med* 348: 538-549.
4. Goldstein JA (2001) Clinical relevance of genetic polymorphisms in the human CYP2C subfamily. *Br J Clin Pharmacol* 52: 349-355.
5. Kupfer A, Desmond PV, Schenker SBR (1979) Family study of a genetically determined deficiency of mephenytoin hydroxylation in man (letter). *Pharmacologist* 21: 173.
6. Wrighton SA, Stevens JC, Becker GW, VandenBranden M (1993) Isolation and characterization of human liver cytochrome P450 2C19: correlation between 2C19 and S-mephenytoin 4'-hydroxylation. *Arch Biochem Biophys* 306: 240-245.
7. Goldstein JA, de Morais SM (1994) Biochemistry and molecular biology of the human CYP2C subfamily. *Pharmacogenetics* 4: 285-299.
8. de Morais SM, Wilkinson GR, Blaisdell J, Nakamura K, Meyer UA, et al. (1994) The major genetic defect responsible for the polymorphism of S-mephenytoin metabolism in humans. *J Biol Chem* 269: 15419-15422.
9. Chang M, Dahl ML, Tybring G, Götharson E, Bertilsson L (1995) Use of omeprazole as a probe drug for CYP2C19 phenotype in Swedish Caucasians: comparison with S-mephenytoin hydroxylation phenotype and CYP2C19 genotype. *Pharmacogenetics* 5: 358-363.
10. De Morais SM, Wilkinson GR, Blaisdell J, Meyer UA, Nakamura K, et al. (1994) Identification of a new genetic defect responsible for the polymorphism of (S)-mephenytoin metabolism in Japanese. *Mol Pharmacol* 46: 594-598.
11. Bråsen K, de Morais SM, Meyer UA, Goldstein JA (1995) A multifamily study on the relationship between CYP2C19 genotype and s-mephenytoin oxidation phenotype. *Pharmacogenetics* 5: 312-317.
12. Mega JL, Close SL, Wiviott SD, Shen L, Hockett RD, et al. (2009) Cytochrome p-450 polymorphisms and response to clopidogrel. *N Engl J Med* 360: 354-362.
13. Luo HR, Poland RE, Lin KM, Wan YJ (2006) Genetic polymorphism of cytochrome P450 2C19 in Mexican Americans: a cross-ethnic comparative study. *Clin Pharmacol Ther* 80: 33-40.
14. Giusti B, Gori AM, Marcucci R, Saracini C, Sestini I, et al. (2009) Relation of cytochrome P450 2C19 loss-of-function polymorphism to occurrence of drug-eluting coronary stent thrombosis. *Am J Cardiol* 103: 806-811.
15. Bonello L, Armero S, Ait Mokhtar O, Mancini J, Aldebert P, et al. (2010) Clopidogrel loading dose adjustment according to platelet reactivity monitoring in patients carrying the 2C19*2 loss of function polymorphism. *J Am Coll Cardiol* 56: 1630-1636.
16. Collet JP, Hulot JS, Pena A, Villard E, Esteve JB, et al. (2009) Cytochrome P450 2C19 polymorphism in young patients treated with clopidogrel after myocardial infarction: a cohort study. *Lancet* 373: 309-317.
17. Geisler T, Schaeffeler E, Dippon J, Winter S, Buse V, et al. (2008) CYP2C19 and nongenetic factors predict poor responsiveness to clopidogrel loading dose after coronary stent implantation. *Pharmacogenomics* 9: 1251-1259.
18. Pedersen RS, Brasch-Andersen C, Sim SC, Bergmann TK, Halling J, et al. (2010) Linkage disequilibrium between the CYP2C19*17 allele and wildtype CYP2C8 and CYP2C9 alleles: identification of CYP2C haplotypes in healthy Nordic populations. *Eur J Clin Pharmacol* 66: 1199-1205.
19. Ramsjö M, Aklillu E, Bohman L, Ingelman-Sundberg M, Roh HK, et al. (2010) CYP2C19 activity comparison between Swedes and Koreans: effect of genotype, sex, oral contraceptive use, and smoking. *Eur J Clin Pharmacol* 66: 871-877.
20. Sim SC, Risinger C, Dahl ML, Aklillu E, Christensen M, et al. (2006) A common novel CYP2C19 gene variant causes ultrarapid drug metabolism relevant for the drug response to proton pump inhibitors and antidepressants. *Clin Pharmacol Ther* 79: 103-113.
21. Sugimoto K, Uno T, Yamazaki H, Tateishi T (2008) Limited frequency of the CYP2C19*17 allele and its minor role in a Japanese population. *Br J Clin Pharmacol* 65: 437-439.
22. Sohn DR, Kusaka M, Ishizaki T, Shin SG, Jang IJ, et al. (1992) Incidence of S-mephenytoin hydroxylation deficiency in a Korean population and the interphenotypic differences in diazepam pharmacokinetics. *Clin Pharmacol Ther* 52: 160-169.
23. Roh HK, Dahl ML, Tybring G, Yamada H, Cha YN, et al. (1996) CYP2C19 genotype and phenotype determined by omeprazole in a Korean population. *Pharmacogenetics* 6: 547-551.
24. Nakamura K, Goto F, Ray WA, McAllister CB, Jacqz E, et al. (1985) Interethnic differences in genetic polymorphism of debrisoquin and mephenytoin hydroxylation between Japanese and Caucasian populations. *Clin Pharmacol Ther* 38: 402-408.
25. Jurima M, Inaba T, Kadar D, Kalow W (1985) Genetic polymorphism of mephenytoin p(4')-hydroxylation: difference between Orientals and Caucasians. *Br J Clin Pharmacol* 19: 483-487.
26. Bertilsson L, Lou YQ, Du YL, Liu Y, Kuang TY, et al. (1992) Pronounced differences between native Chinese and Swedish populations in the polymorphic hydroxylations of debrisoquin and S-mephenytoin. *Clin Pharmacol Ther* 51: 388-397.
27. Jaja C, Burke W, Thummel K, Edwards K, Veenstra DL (2008) Cytochrome p450 enzyme polymorphism frequency in indigenous and native american populations: a systematic review. *Community Genet* 11: 141-149.
28. Freidin MB, Bragina E Yu, Petruskiy FI (2003) Association of the GSTT1, GSTM1, CYP2C19, CYP2E1 Genes polymorphism with atopy. *Med Immunol* 1-2: 107-112.
29. Bonello L, Armero S, Ait Mokhtar O, Mancini J, Aldebert P, et al. (2010) Clopidogrel loading dose adjustment according to platelet reactivity monitoring in patients carrying the 2C19*2 loss of function polymorphism. *J Am Coll Cardiol* 56: 1630-1636.
30. Kantemirova BI, Timofeeva NV, Sychev DA (2011) A comparative study of CYP2C19 gene polymorphism in children living in the Astrakhanian region. *Achievement Of Science In Practice* 2: 136-143.
31. Romodanovsky DP, Khapaev BA, Ignatiev V, Kukes VG, Karkishchenko VN, et al. (2010) Frequency "slow" allele variants of the genes coding isoenzymes of cytochrome P450: CYP2D6, CYP2C19, CYP2C9 in Karachaevs and Circassians. *Biomedicine* 2: 33-37.
32. Mirzaev KB, Mammaev SN, Gafurov DM, Kazakov RE, Sychev DA (2014) Prevalence of polymorphic markers of CYP2C19 * 2 (G681A, rs4244285) and significance for personalization pharmacotherapy population mountaineers of Dagestan. *Russian medical lead: In press*.
33. Nizhevich AA, lunusbaev BB, TuĀgunov MM, Tsiglintseva NP, Nasretdinova EK (2009) [Study of gene polymorphism responsible for metabolism of proton pump inhibitors in children with H. pylori infection: is there a correlation with efficacy of eradication treatment?]. *Eksp Klin Gastroenterol* : 101-104.
34. Makeeva O, Stepanov V, Puzyrev V, Goldstein DB, Grossman I (2008) Global pharmacogenetics: genetic substructure of Eurasian populations and its effect on variants of drug-metabolizing enzymes. *Pharmacogenomics* 9: 847-868.
35. Mirzaev KB, Sychev DA, Karkishchenko VN (2013) CYP2C19*2, CYP2C19*3, CYP2C19*17 Allele and Genotype Frequencies in Clopidogrel-Treated Patients with Coronary Heart Disease from the Russian Population. *Biomedicine* 2013; 1:117-128.
36. Komarov AL, Panchenko EP, Donnikov AE, Shakhmatova OO, Dzhalilova GV, et al. (2011) [Factors determining clinical effectiveness of clopidogrel and prognosis of patients with stable ischemic heart disease]. *Kardiologija* 51: 8-18.
37. Gorbachenko EA (2011) Genetic polymorphism of enzymes of xenobiotic metabolism and the formation of resistance to chemotherapy in patients with chronic lymphoproliferative disorders. *Materials XLIX International Scientific Student Conference "Student and scientific and technological progress", Novosibirsk, Russian Federation*.
38. Gaikovitch EA, Cascorbi I, Mrozkiewicz PM, Brockmüller J, Frötschl R, et al. (2003) Polymorphisms of drug-metabolizing enzymes CYP2C9, CYP2C19, CYP2D6, CYP1A, NAT2 and of P-glycoprotein in a Russian population. *Eur J Clin Pharmacol* 59: 303-312.
39. Galiavich AS, Valeeva DD, Minnetdinov RSh, Arkhipova AA, Akhmetov II, et al. (2012) [CYP2C19 gene polymorphism in patients with myocardial infarction who use clopidogrel]. *Kardiologija* 52: 20-24.
40. Khalikova AR, Arkhipova AA Akhmetov II (2012) The study of cytochrome P-450 CYP2C19 gene polymorphisms in population of Tatars living in Republic of Tatarstan. *Prakticheskaja Medicina* 3: 53-55.

Citation: Mirzaev K, Sychev D, Arutyunyan G, Yugay A, Andreev D (2014) Cytochrome 2C19 Enzyme Polymorphism Frequency in Different Indigenous Ethnic Groups in Russian Federation: A Systematic Review *J Pharmacogenomics Pharmacoproteomics* 5: 136. doi:10.4172/2153-0645.1000136